



# The effects of dietary carbohydrate level on the growth performance, body composition and feed utilization of juvenile Kelabau (*Osteochilus melanopleurus*)

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**Abstract.** This study was conducted to evaluate the effect of feeding with different carbohydrate (CHO) content on growth performance and body composition in kelabau (*Osteochilus melanopleurus*). Treatments consisted in 3 experimental iso-nitrogenous and iso-lipid diets, at different CHO levels: 30.50% (A), 32.76% (B), and 34.83% (C). Treatments were applied to *O. melanopleurus* culture specimens, with an average weight of  $3.18 \pm 0.26$  g and a stocking density of 20 individuals, maintained in plastic tubs measuring 54.3 x 38 x 31.5 cm and filled with 40 L of water. *O. melanopleurus* was fed 2 times a day at satiation basis, for 60 days. The result showed that fish fed with diet B (32.76%) had higher growth performance and feed utilization compared to the other groups (A and C) ( $P < 0.05$ ). The increase in carbohydrate administration in *O. melanopleurus* from 30.50% to 34.82% CHO does not affect the liver and muscle glycogen levels, but does affect the lipid liver content.

**Key Words:** herbivorous fish, freshwater, lipid content, protein retention, energy retention.

**Introduction.** Indonesian freshwater is rich in herbivorous fish species, especially the Cyprinidae family, including the kelabau, *Osteochilus melanopleurus* (Bleeker). In 2013, *O. melanopleurus* began to be cultivated until nowadays. This fish ecologically plays an important role in river and lake ecosystems, because they are classified as herbivorous fish. In the digestive tract of *O. melanopleurus*, several species of plants and phytoplankton and algae are found. In young fish of a size between 200 and 299 mm, the intestine contains 83.3% plants, whereas in larger fish (300 mm) the intestine content is 100% plants (Aizam et al 1983). Fish that consume feed from complex sources with the carbohydrate content of 31% produce the best relative growth (RGR), reaching 49.45%, compared to other treatments (Mardani 2014). Susanto et al (2019) reported that *O. melanopleurus* that consumed feed with a protein content of 31.88% and a carbohydrate content of 30.53% gave the best growth.

Fish have a lower ability to utilize carbohydrates than land animals, but carbohydrates must be available in fish feed, because if carbohydrates are not available then other nutrients such as protein and lipid will be metabolized to be used as energy so that fish growth will be slow (Wilson 1994). Yamada (1983) further explained that the carbohydrates concentrations in fish feed depend on the complexity of the carbohydrates. Carnivorous fish are unable to utilize complex carbohydrates in their feed at high levels. However carnivorous fish can utilize simple carbohydrates such as glucose, sucrose, and lactose as the main energy source. Furthermore, Furuichi (1988) stated that carnivorous fish can optimize carbohydrates use at the level of 10-20% in their feed and omnivorous

fish reaches the optimal efficiency at the level of 30-40% carbohydrates in their feed. The results of the Seenappa & Devaraj (1995) experiments, using three levels of carbohydrates (15, 25 and 35%) on Indian major carps *Catla catla*, showed that the best fish growth was recorded at 35% carbohydrates.

Freshwater fish and sea fish have different abilities in digesting carbohydrates. The ability of sea fish to digest carbohydrates is around 20%, while freshwater fish reach 30-40%, in the case of *Cyprinus carpio* (Satoh 1991), 25-30% in the case of *Ictalurus punctatus* (Wilson 1991) and about 40% in the case of *Tilapia* sp. (Luquet 1991). Hernandez et al (1995) observed that the administration of carbohydrates to tambaqui (*Colossoma macropomum*) juveniles at a weight of 0.5 g is as effective as lipids, in terms of energy sources. Gunther (1996) further stated that *C. macropomum* can efficiently use carbohydrates and produce the best growth by feeding with the carbohydrate content of 38%, whereas, in grass carp (*Ctenopharyngodon idella*), the optimal growth occurs in fish that consume feed at a 27.5% carbohydrate level (Gau et al 2010). In roho labeo (*Labeo rohita*), which consumes feed with an increase in carbohydrates from 30% to 40% along with a decrease in protein content from 40% to 30%, an increase in Protein Efficiency Ratio (PER) is observed (Erfanullah & Jafri 1995). Research on the carbohydrate requirements of *O. melanopleurus* has not yet been conducted, which is the rationale for the current research aiming to determine the carbohydrate levels corresponding to an optimal growth of *O. melanopleurus*.

## Material and Method

**Diets.** This study used 3 kinds of artificial feed which are isonitrogenous (32.1%) and isolipid (9.1%) with different carbohydrate content, namely feed A (30.5%), feed B (32.7%), and C feed (34.8%) with a calorie to protein (CP) ratio ranging from 8.0 to 8.3 kcal. Feed formulations can be seen in Table 1.

Table 1  
Treatment feed composition (g) and feed nutrient content based on dry weight

Ingredients	Ingredient percentage in the trial feed (% dry matter)		
	A (30.50% CHO)	B (32.76% CHO)	C (34.82% CHO)
Fish meal	29.1	28.7	28.7
Soybean meal	25.8	25.7	25.2
Wheat meal	13.0	15.3	17.9
Brand meal	9.0	9.2	9.1
Fish oil	2.5	2.5	2.5
Corn oil	2.5	2.5	2.5
Vitamin mix*	3.0	3.0	3.0
Mineral mix**	3.0	3.0	3.0
Choline chloride	2.0	2.0	2.0
CMC	2.0	2.0	2.0
Filler	8.1	6.1	4.1
Proximate analysis result			
Protein (%)	31.26	31.38	31.29
NFE (%)	30.50	32.76	34.82
Lipid (%)	9.11	9.54	9.57
Fiber (%)	15.55	12.61	10.59
Total energy (Kcal g <sup>-1</sup> )***	259.46	269.01	274.13
E/P (Kcal g <sup>-1</sup> protein)	8.30	8.57	8.76

CMC-carboxymethyl cellulose; NFE-nitrogen free extract; E/P-energy protein ratio; \*in mg kg<sup>-1</sup> feed: vit. B<sub>1</sub> 60; vit. B<sub>2</sub> 100; vit. B<sub>12</sub> 100; vit. C 2000; vit. K<sub>3</sub> 50; vit. A/D<sub>3</sub>400; \*\*in mg kg<sup>-1</sup> feed: MgSO<sub>4</sub>.7H<sub>2</sub>O 7.5; NaCl 0.5; NaH<sub>2</sub>PO<sub>4</sub>.2H<sub>2</sub>O 12.5; KH<sub>2</sub>PO<sub>4</sub> 16.0; CaHPO<sub>4</sub>.2H<sub>2</sub>O 6.53; Fe citric 1.25; ZnSO<sub>4</sub>.7H<sub>2</sub>O 0.1765; MnSO<sub>4</sub>.4H<sub>2</sub>O 0.081; CuSO<sub>4</sub>.5H<sub>2</sub>O 0.0155; KIO<sub>3</sub> 0.0015; CoSO<sub>4</sub> 0.0003; \*\*\*protein=3.5 kcal g<sup>-1</sup>; NFE=2.5 kcal g<sup>-1</sup>; lipid=8.1 kcal g<sup>-1</sup>.

**Fish culture management.** *O. melanopleurus* specimens were obtained from the hatchery of a Freshwater Aquaculture Center (Balai Benih Air Tawar), in Mandiangin,

South Kalimantan. Fish were reared in a plastic tank containing 40 L water with a density of 20 fish per tank with an average weight of  $3.18 \pm 0.26$  g. Fish were cultured for 60 days in a semi-closed circulation system, being fed twice a day in the morning and evening, at satiation. Dejections were collected in the morning. Water lost due to the siphon was replaced by new water up to the same volume. The filter was washed every day and the filter bath was washed and replaced with new water every week. During the study, the average water temperature was  $28.5 \pm 1.0^\circ\text{C}$ , dissolved oxygen was 4.50-6.10  $\text{mg L}^{-1}$ , pH was between 6.80 and 6.95, total ammonia nitrogen was between 0.382 and 0.623  $\text{mg L}^{-1}$ . This shows that the water conditions during the study were optimal (Tebbut 1992; Effendie 1997).

**Data collection and chemical analysis.** Initial and final body weights were measured upon anesthetized fish (using MS 222). Weighing was performed daily to determine the specific growth rate (SGR) according to De Silva & Anderson (1995). During the study, the following data were recorded: feed consumed, to determine the Total Feed Consumption (TFC), according to Pereira et al (2007); feed efficiency, protein retention, lipid retention and energy retention, according to NRC (2011); Protein Efficiency Ratio (PER), according to Bake et al (2014). Proximate body analysis was carried out at the beginning and at the end of the study, to determine the nutrient composition of fish (Takeuchi 1988). Analysis of liver and meat glycogen, as well as liver lipid content, was carried out at the beginning and end of the study to determine the reserve energy (Takeuchi 1988).

**Statistic analysis.** The design of this study followed an experimental laboratory model, using a completely randomized design (CRD) consisting of three treatments and five replications. The data of feed efficiency (FE), weight growth, total feed consumption (TFC), protein retention (PR), lipid retention (LR), energy retention (ER) and protein efficiency ratio (PER) were tested for variance with ANOVA, followed by a Tukey test at 95% confidence interval, using the SPSS program, while liver and muscle glycogen levels, as well as liver lipid levels, were analyzed descriptively in tabular form.

## Results and Discussion

**Growth performance and feed utilization efficiency.** The values of various parameters of feed use, including: weight gain, specific growth rate, relative growth rate, protein retention, lipid retention, energy retention and the ratio of protein efficiency. as well as feed efficiency, were determined for a 60 days rearing period and three feed treatments with different carbohydrate-concentrations, as presented in Table 2.

The change in the carbohydrate concentration levels have a significant influence on the weight growth, relative growth rate, specific growth rate, protein efficiency ratio (PER), feed efficiency, protein retention, lipid retention and energy retention ( $P < 0.05$ ), and did not significantly affect the level of feed consumption ( $P > 0.05$ ). The best weight growth was obtained in the treatment B (32.76% CHO), followed by C (34.82% CHO). The lowest weight growth was obtained in the group of fish that consumed feed A (30.50% CHO) ( $P < 0.05$ ).

The best relative growth rate was also obtained in the group of fish fed with B (32.76% CHO) which was 0.84% per day statistically higher than in fish that consumed other feed ( $P < 0.05$ ). The same phenomenon was also seen in the specific growth rate, where the group of fish that consumed feed B had the best specific growth rate of 1.04% per day statistically higher than the group of fish that consumed feed C and A ( $P < 0.05$ ).

Table 2

The average value of initial weight, final weight, growth weight, relative growth rate (RGR), specific growth rate (SGR), total feed consumption (TFC), protein efficiency ratio (PER), feed efficiency (FE), protein retention (PR), lipid retention (LR), energy retention (ER) and total ammonia nitrogen (TAN) obtained in *Osteochilus melanopleurus* maintained for 60 days by feeding with different carbohydrates

Parameters	CHO (%)		
	A(30.50)	B(32.76)	C(34.82)
Initial weight (g)	3.17±0.32 <sup>a</sup>	3.08±0.23 <sup>a</sup>	3.24±0.29 <sup>a</sup>
Final weight (g)	4.86±0.14 <sup>a</sup>	5.65±0.09 <sup>c</sup>	5.14±0.06 <sup>b</sup>
Weight growth (g)	33.71±5.14 <sup>a</sup>	51.39±5.49 <sup>b</sup>	37.48±4.90 <sup>a</sup>
RGR (%)	0.54±0.12 <sup>a</sup>	0.84±0.14 <sup>b</sup>	0.60±0.15 <sup>a</sup>
SGR (%)	0.72±0.14 <sup>a</sup>	1.04±0.08 <sup>b</sup>	0.80±0.06 <sup>a</sup>
TFC (%)	121.26±4.57 <sup>a</sup>	127.52±3.82 <sup>a</sup>	125.89±3.16 <sup>a</sup>
PER (%)	0.89±0.12 <sup>a</sup>	1.28±0.11 <sup>b</sup>	0.95±0.13 <sup>a</sup>
FE (%)	27.74±3.69 <sup>a</sup>	40.23±3.34 <sup>b</sup>	29.80±4.13 <sup>a</sup>
PR (%)	61.64±7.33 <sup>a</sup>	84.17±7.91 <sup>b</sup>	66.19±8.05 <sup>a</sup>
LR (%)	62.51±4.51 <sup>a</sup>	91.06±8.09 <sup>b</sup>	76.41±13.19 <sup>ab</sup>
ER (%)	45.00±4.13 <sup>a</sup>	63.34±3.45 <sup>b</sup>	49.40±7.01 <sup>a</sup>
TAN (mg (g body hour) <sup>-1</sup> )	0.00170±0.00004 <sup>c</sup>	0.00124±0.00007 <sup>a</sup>	0.00154±0.00012 <sup>b</sup>

The average followed by different superscript letters in the same row shows significant difference (P<0.05).

*O. melanopleurus* that was maintained by feeding B showed the same feed consumption levels as the fish fed with C and A (P>0.05). Fish that consumed feed B have the highest protein efficiency ratio (PER), compared to fish that consumed feed A and C (P<0.05), which is 1.28%. The best value of feed efficiency was obtained in fish that consumed feed B, which was 40.23±3.34%, higher than in the group of fish that consumed feed C, with a value of feed efficiency of 29.80±4.13%, while fish that consumed feed A produced the lowest feed efficiency, at a level of 27.74±3.69%.

Fish that consumed feed B had the highest protein retention value (84.17±7.91%), compared to the specimens that consumed feed A and C (66.19±8.05% and 61.64±7.33%, respectively). The lipid retention value in fish that consumed feed B was also the highest, with a value of 91.06±8.09%, followed by the group of fish that consumed feed C and A. The same observation stands for the energy retention value in the group of fish that consumed feed B, which was higher, with a value of 63.34±3.45%, than in specimens that consumed feed A (45.00±4.13%). The group of fish that consumed feed B produced a TAN excretion of 0.00124±0.00007 mg (g body hour)<sup>-1</sup>, lower than the group of fish that consumed feed C, followed by group A (P<0.05).

**Initial and final proximate analysis, liver and muscle glycogen content, and liver lipid content.** Initial and final proximate composition, after being reared for 60 days by feeding with different carbohydrate contents, is presented in Table 3.

The protein level of the fish body at the end of the study decreased, but it was not influenced by the increase in feed carbohydrates (to the level of 34.82% CHO) and had no significant effect on fish body protein levels (P>0.05). The same phenomenon was also found in the ash content of the fish body at the end of the study, but the administration of different carbohydrate levels significantly affected the ash content (P<0.05). At the opposite, increasing the carbohydrate levels does not have a significant effect on the NFE content (P>0.05). The increase in feed carbohydrates significantly affected the fish body lipid content, the highest value being obtained in the group of fish that consumed feed with 32.76% CHO (P<0.05).

Table 3

Initial and final proximate body composition of *Osteochilus melanopleurus* maintained for 60 days by feeding with different carbohydrates (% dry weight)

Parameters	Level CHO (%)					
	A (30.50)		B (32.76)		C (34.82)	
<i>Initial body composition (%)</i>						
Protein	62.41		62.41		62.41	
Lipid	5.87		5.87		5.87	
Ash	21.04		21.04		21.04	
NFE	9.43		9.43		9.43	
<i>Final body composition (%)</i>						
Protein	64.86	±0.62 <sup>a</sup>	63.87	±1.13 <sup>a</sup>	64.64	±1.11 <sup>a</sup>
Lipid 01	10.94	±0.49 <sup>a</sup>	13.01	±0.95 <sup>b</sup>	12.63	±1.17 <sup>b</sup>
Ash	15.11	±0.46 <sup>b</sup>	13.62	±0.46 <sup>a</sup>	14.29	±0.55 <sup>ab</sup>
NFE	7.77	±0.55 <sup>a</sup>	8.57	±0.59 <sup>a</sup>	7.73	±0.47 <sup>a</sup>

The average followed by different superscript letters in the same row shows significant difference (P<0.05).

Liver and muscle glycogen levels, as well as initial and final liver lipid content in *O. melanopleurus* maintained for 60 days by feeding with different carbohydrates, are presented in Table 4.

Maintenance of fish for 60 days with feed containing different carbohydrates does not have a significant effect on the liver and muscle glycogen levels. Liver glycogen levels of fish fed with C (34.82% CHO) had liver glycogen levels of 2.14±0.11 µg g<sup>-1</sup> which were relatively similar compared to fish consuming B and A feeds with liver glycogen levels of 2.11±0.10 µg g<sup>-1</sup> and 2.09±0.13 µg g<sup>-1</sup>, respectively. Glycogen levels of fish muscle fed with C (34.82% CHO), reaching 0.178±0.009 µg g<sup>-1</sup>, were the same as in fish that consumed feed B and C with muscle glycogen levels of 0.176±0.008 µg g<sup>-1</sup> and 0.174±0.010 µg g<sup>-1</sup>, respectively.

The highest level of liver lipid was found in the group of fish that consumed feed B, reaching 6.50±0.47%, followed by fish that consumed feed C and feed A with liver lipid contents of 6.31±0.58% and 5.47±0.25%, respectively.

Table 4

Levels of the liver and muscle glycogen and liver lipid content *Osteochilus melanopleurus* were maintained for 60 days by giving feed containing different carbohydrates

Parameters	Levels CHO (%)					
	A (30.50)		B (32.76)		C (34.82)	
<i>Glycogen content (µg g<sup>-1</sup>)</i>						
Liver	2.09	±0.13 <sup>a</sup>	2.11	±0.10 <sup>a</sup>	2.14	±0.11 <sup>a</sup>
Muscle	0.174	±0.010 <sup>a</sup>	0.176	±0.008 <sup>a</sup>	0.178	±0.009 <sup>a</sup>
<i>Liver lipid content (%)</i>						
Initial	2.94		2.94		2.94	
Final	5.47±0.25 <sup>a</sup>		6.50±0.47 <sup>b</sup>		6.31±0.58 <sup>b</sup>	

The average followed by different superscript letters in the same row shows significant difference (P<0.05).

**Discussion.** Feeding with an increase in carbohydrate levels up to 32.76% significantly increases the observed specimens' growth performance. Conversely, the latter decreases by rising the carbohydrate content up to 34.82%. This indicates that in the treatment B scenario (32.76% CHO), non-protein feed energy sources are better processed and more efficient in growth, compared treatments C (34.82% CHO) and A (30.50% CHO).

Bray & Lawrence (1992), states that the availability of energy mainly from carbohydrates as an energy source other than lipids and proteins is mainly used for metabolism, both for growth and subsequently for reproduction in nature. Therefore, if the energy needed for metabolism and growth is sufficient, the exceeding nutrients or energy will be stored or used for reproduction. Wilson (1994) added that if carbohydrates

are deficient, then other nutrients such as protein and lipids will be metabolized to energy, therefore fish growth will be slower.

Feed efficiency and protein efficiency ratio are the determinant parameters for evaluating the effectiveness of feed in growth. The fish group that consumed feed B (32.76% CHO) had a higher feed efficiency and protein efficiency ratio than the other groups. This indicates that the fish can utilize the nutrients they consume, especially carbohydrates and lipids as a source of energy for protein synthesizing and for growth. The results of this study also illustrate the importance of the presence of carbohydrates in the feed. The importance of providing carbohydrates at certain levels was also reported by Castro et al (2016), in gilthead seabream (*Sparus aurata*) juveniles. Fish fed with 20% carbohydrates and different lipid concentrations did not experience differences in growth but higher PER and lipid retention were observed in the group of fish that consumed feed with carbohydrates. Conversely, fish that are fed without carbohydrates, experience lower lipid retention.

Retention ratio describes the stored nutrients fraction of the total intake of nutrients of the same type, during a reference period. The results of this study indicate that the fish group fed B (32.76% CHO), produced Protein Retention (PR), Lipid Retention (LR), and Energy Retention (ER) higher than the C and A fish groups. This illustrates that the consumed nutrients, besides being used for activities are also stored in the muscles as energy reserves. The high value of nutrient retention (PR, LR, and ER), in group B fish also indicates that the consumed feed is balanced, so that the portion of protein for growth is not disrupted.

Conversely, an increase in CHO levels to 34.82% CHO caused a decrease in nutrient retention in *O. melanopleurus*. The low value of nutrient retention in the fish group corresponding to treatment C is closely related to the decreased level of feed consumption. The high energy in the feed causes the fish to limit the amount of feed consumed, thus affecting the number of feed nutrients consumed.

In the fish group corresponding to treatment A (30.50% CHO), the low value of nutrient retention is due to the suboptimal use of the feed, which does not provide enough energy for standard activities, and requires compensation via protein and lipids metabolizing. Mokoginta et al (1995) explained that if the energy content of the feed was too low, then most of the feed protein would be catabolized to meet energy needs, so that fish consumes larger amounts of food. Fish will limit the amount of feed consumption if the feed contains too high energy because basic energy needs have been met. An indicator of the presence of proteins catabolized for energy in both groups of fish corresponding to treatments C and A is the higher level of total ammonia nitrogen (TAN) excretion value of  $0.00170 \pm 0.00004$  mg (g body hour)<sup>-1</sup> and  $0.00154 \pm 0.00012$  mg (g body hour)<sup>-1</sup>, respectively, compared to fish group of treatment B, with only  $0.00124 \pm 0.00007$  mg (g body hour)<sup>-1</sup>.

Carbohydrates administration in higher proportions will inhibit growth. Li et al (2019) stated that the growth performance and utilization of feed decreased in groupers by increasing carbohydrate content of the feed, as seen from glucose intolerance in these fish. The same phenomenon was observed by Zhou et al (2013) in *Megalobrama amblycephala*, by Wu et al (2016) in juvenile black carp *Mylopharyngodon piceus*, by Dong et al (2016) on golden pompano *Trachinotus ovatus*, by Wang et al (2016) in juvenile grouper *Epinephelus akaara* and by Xie et al (2017) in Nile tilapia *Oreochromis niloticus*. Ren et al (2011) also stated that increasing levels of carbohydrate to 18.4% CHO for juvenile cobia *Rachycentron canadum* will increase SGR, FER, and PER.

The provision of carbohydrate feed, which is increasing in fish from 30.50% CHO to 34.82% CHO, does not affect the level of liver glycogen and muscle but does affect the level of liver lipid. Similar results were obtained by Wang et al (2016), who stated that an increase in feed carbohydrates will lead to the accumulation of liver glycogen in carnivorous fish such as juvenile groupers *E. akaara* and large yellow croaker juveniles *Larimichthys crocea* (Xing et al 2016). Guo et al (2015) stated that grass carp *C. idella* herbivorous fish, fed with carbohydrate content increased from 30.94% to 42.31%, did not experience alterations of the growth performance, but only an increased level of liver lipid and liver glycogen. In omnivores such as Prussian carp, *Carassius gibelio*, high levels

of carbohydrates with low lipid (45.0% CHO and 2.0% lipid) also did not cause differences in specific growth rates but increased Hepatosomatic Index (HSI), liver lipid content and lipid retention efficiency (Li et al 2019). Similar observations were done by Mozanzadeh et al (2016) on juvenile Sobaity seabream *Sparidentex hasta* fed with increased carbohydrate content and reduced dietary lipid content, which did not influence the growth performance, in particular the weight growth, condition factors, specific growth rate, feed consumption and feed conversion rates.

Based on the growth performance and feed utilization, the optimum level of carbohydrate for *O. melanopleurus* is 32.76%. These results are almost the same as reported by Boonuntanasarn et al (2018) in tilapia *O. niloticus* where the optimal level of carbohydrates is 32.6%. Lower yields have been reported in other fish such as: the silver barb *Puntius gonionotus* (29.3-29.8%) by Mohanta et al (2009); the juvenile cobia *Rachycentron candum* L (21.1% CHO), by Ren et al (2011); the Wuchang Bream *Megalobrama amblycephala* (31.0%), by Zhou et al (2015); the golden pompano *T. ovatus* (16.93-20.64%), by Dong et al (2016); the juvenile large yellow croaker, *L. crocea* (21.29%), by Xing et al (2016) and 16.27% CHO by Zhou et al (2016), and the juvenile black carp *M. piceus* (24.98%), by Wu et al (2016). The optimum level of carbohydrate 32.76% in this study was also higher than the optimum value obtained by Xie et al (2017) in juvenile tilapia *O. niloticus* (28.87%), by Asemani et al (2019) in striped catfish *Pangasianodon hypophthalmus* fingerlings (30.81-31.13%) and by Yanto et al (2019) in *Barbonymus schwanenfeldii* (22.89 g%).

Higher optimal carbohydrate levels in feed were found by Seenappa & Devaraj (1995) in major carp *Catla catla* (35% CHO), by Luquet (1991) in *Tilapia* sp. (40.0% CHO) and by Mokoginta et al (2004) in gourami *Osphronemus goramy* measuring 78.7 g (47.5% CHO). Mohapatra et al (2003) reported that roho labeo (*Labeo rohita*) that consumed feed with a carbohydrate content of 45.0% at a protein content of 30% was more efficient in using the feed.

Based on the results of the research above, further research is necessary to discover the optimal carbohydrate and lipid ratio so that the growth of *O. melanopleurus* can be maximized.

**Conclusions.** *O. melanopleurus* that consume 32.76% CHO provided the best weight growth, relative growth, feed efficiency, protein efficiency ratio, protein retention, lipid retention and energy retention compared to other treatments. Increasing carbohydrate levels up to 34.82% did not increase liver and muscle glycogen levels and liver lipid content.

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## References

- Aizam Z. A., Roos S. C., Ang K. J., 1983 Some aspects of the Biology of Kelabau *Osteochilus melanopleurus* (Bleeker). *Pertanika* 6(3):99-106.
- Asemani M., Sepahdari A., Pourkazemi M., Hafezieh M., Aliyu-Paiko M., Dadgar S., 2019 Effect of different sources and form of dietary carbohydrates on growth performance, body indices, and lipogenesis activity of striped catfish *Pangasianodon hypophthalmus* fingerlings. *Aquaculture Nutrition* 25(6):1399-1409.
- Bake G. G., Martins E. I., Sadiku S. O. E., 2014 Nutritional evaluation of varying levels of cooked flamboyant seed meal (*Delonix regia*) on the growth performance and body composition of Nile tilapia (*Oreochromis niloticus*) fingerlings. *Agriculture, Forestry, and Fisheries* 3(4):233-239.
- Boonuntanasarn S., Jangprai A., Kumkhong S., Juan P. E., Veron V., Burel C., Marandel L., Pansera S., 2018 Adaptation of Nile tilapia (*Oreochromis niloticus*) to different

- levels of dietary carbohydrates: New insights from a long-term nutritional study. *Aquaculture* 496:58-65.
- Bray W. A., Lawrence A. L., 1992 Reproduction of *Penaeus* species in captivity. In: Marine shrimp culture: principles and practices. Fast A. W., Lester J. L. (eds), Elsevier, Amsterdam, 862 p.
- Castro C., Corraze G., Firmino-Diógenes A., Larroquet L., Panserat S., Oliva-Teles A., 2016 Regulation of glucose and lipid metabolism by dietary carbohydrate levels and lipid sources in gilthead seabream juveniles. *British Journal of Nutrition* 116(1):19-34.
- De Silva S. S., Anderson T., 1995 Fish nutrition in aquaculture. Springer Science & Business Media, 320 p.
- Dong L. F., Tong T., Zang Q., Wang Q. C., Xu M. Z., Yu H. R., Wang J., 2016 Effect of dietary carbohydrate to lipid ratio on growth, feed utilization, body composition, and digestive enzyme activities of golden pompano (*Trachinotus ovatus*). *Aquaculture Nutrition* 24(1):341-347.
- Effendie M. I., 1997 [Fisheries biology methods]. Yayasan Pustaka Nusantara, Yogyakarta, Indonesia, 258 p. [In Indonesian].
- Erfanullah, Jafri A. K., 1995 Protein-sparing effect of dietary carbohydrate in diets for fingerling *Labeo rohita*. *Aquaculture* 136:331-339.
- Furuichi M., 1988 Dietary requirements. In: Fish nutrition and mariculture. Watanabe T. (ed), Kanazawa International Fisheries Centre, Japan International Corporation Centre, pp. 21-78.
- Gau W., Liu Y. J., Tian L. X., Mai K. S., Liang G. Y., Yang H. J., Huai M. Y., Luo W. J., 2010 Effect of dietary carbohydrate-to-lipid ratios on growth performance, body composition, nutrient utilization and hepatic enzymes activities of herbivorous grass carp (*Ctenopharyngodon idella*). *Aquaculture Nutrition* 16:327-333.
- Gunther N. J., 1996 Growth of tambaqui (*Colossoma macropomum*) juveniles at different carbohydrate-lipid ratios. *Journal of Aquaculture in the Tropics* 11(2):105-112.
- Guo X., Liang X. F., Fang L., Yuan X., Zhou Y., Zhang J, Li B., 2015 Effects of dietary non-protein energy source levels on growth performance, body composition, and lipid metabolism in herbivorous grass carp (*Ctenopharyngodon idella* Val.). *Aquaculture Research* 46:1197-1208.
- Hernández M., Takeuchi T., Watanabe T., 1995 Effect of dietary energy sources on the utilization of protein by *Colossoma macropomum* fingerlings. *Fisheries Science* 61:507-511.
- Li H., Xu W., Jin J., Zhu X., Yang Y., Han D., Liu H, Xie S., 2019 Effect of dietary carbohydrate and lipid concentrations on growth performance, feed utilization, glucose, and lipid metabolism in two strains gibel carp (*Carrasius auratus gibelio*). *Frontiers in Veterinary Science* 6:165. doi: 10.3389/fvets.2019.00165.
- Li S., Li Z., Zhang J., Sang C., Chen N., 2019 The impacts of dietary carbohydrate levels on growth performance, feed utilization, glycogen accumulation, and hepatic glucose metabolism in hybrid grouper (*Epinephelus fuscoguttatus* ♀ × *E. lanceolatus* ♂). *Aquaculture* 512:734351.
- Luquet P., 1991 Tilapia, *Oreochromis* spp. In: Handbook of nutrient requirements of finfish. Wilson R. P. (ed), pp. 169-179, CRC Press, Boca Raton, FL.
- Mardani, 2014 [The effect of different food sources on the growth of Kelabau Padi *Osteochilus melanopleura* maintained in Hapa in ponds]. *Jurnal Ilmu Hewani Tropika* 3(1):22-26. [In Indonesian].
- Mohanta K. N., Mohanty S. N., Jena J., Sahu N. P., Patro B., 2009 Carbohydrate level in the diet of silver barb *Puntius gonionotus* (Bleeker) fingerlings: effect on growth, nutrient utilization, and whole-body composition. *Aquaculture Research* 40:927-937.
- Mohapatra M., Sahu N. P., Chaudari A., 2003 Utilization of gelatinized carbohydrate in diets of *Labeo rohita* fry. *Aquaculture Nutrition* 9:189-196.
- Mokoginta I., Suprayudi M. A., Setiawati M., 1995 [Optimum protein and energy requirement of giant gouramy (*Osphronemus gouramy* Lac.) fry's feed]. *Jurnal Penelitian Perikanan Indonesia* 1(3):82-94. [In Indonesian].



- Mokoginta I., Takeuchi T., Hadadi A., Dedi J., 2004 Different capabilities in utilizing dietary carbohydrates by fingerling and subadult giant gourami *Osphronemus gourami*. *Fisheries Science* 70:996-1002.
- Mozanzadeh M. T., Yavari V., Marammazi J. G., Agh N., Gisbert E., 2016 Optimal dietary carbohydrate to lipid ratio for silvery-black porgy (*Sparidentex hasta*) juveniles. *Aquaculture Nutrition* 24(1):341-347.
- Pereira L., Requelme T., Hosokawa H., 2007 Effect of three photoperiod regimes on the growth and mortality of Javanese abalone *Helios discus* (Hanoino). *Journal of Shellfish Research* 26:763-767.
- Ren M., Ai Q., Mai K., Ma H., Wang X., 2011 Effect of dietary carbohydrate level on growth performance, body composition, apparent digestibility coefficient, and digestive enzyme activities of juvenile Cobia, *Rachycentron canadum*. *Aquaculture Research* 42:1467-1475.
- Satoh S., 1991 Common carp *Cyprinus carpio*. In: Handbook of nutrient requirements of finfish. Wilson R. P. (ed), pp. 55-67, CRC Press, Boca Raton, FL.
- Seenappa D., Devaraj K. V., 1995 Effect of different levels of protein, fat, and carbohydrate on growth, feed utilization, and body carcass composition of fingerlings in *Catla catla* (Ham.). *Aquaculture* 129:243-249.
- Susanto A., Hutabarat J., Anggoro S., Subandiyono, 2019 The effects of dietary protein level on the growth, protein efficiency ratio, and body composition of juvenile kelabau (*Osteochilus melanopleurus*). *AAFL Bioflux* 12(1):320-326.
- Takeuchi T., 1988 Laboratory work chemical evaluation of dietary nutrients. In: Fish nutrition and mariculture. Watanabe T. (ed), pp. 179-225, Department of Aquatic Bioscience, Tokyo University of Fisheries, JICA.
- Tebbut T. H. Y., 1992 Principles of water quality control. Fourth edition, Pergamon Press, Oxford, 251 p.
- Wang J., Li X., Han T., Yang Y., Jiang Y., Yang M., Xu Y., Harpaz S., 2016 Effects of different dietary carbohydrate levels on growth, feed utilization, and body composition of juvenile grouper *Epinephelus akaara*. *Aquaculture* 459:143-147.
- Wilson R. P., 1991 Channel catfish *Ictalurus punctatus*. In: Handbook of nutrient requirements of finfish. Wilson R. P. (ed), pp. 35-53, CRC Press, Boca Raton, FL.
- Wilson R. P., 1994 Utilization of dietary carbohydrates by fish. *Aquaculture* 124:67-80.
- Wu C., Ye J., Gao J., Yang X., Zhang Y., 2016 Effect of varying carbohydrate fraction on growth, body composition, metabolic, and hormonal indices in juvenil black carp, *Mylopharyngodon piceus*. *The Journal of World Aquaculture Society* 47(3):435-449.
- Xie D., Yang L., Yu R., Chen F., Lu R., Qin C., Nie G., 2017 Effects of dietary carbohydrate and lipid levels on growth and hepatic lipid deposition of juvenile tilapia, *Oreochromis niloticus*. *Aquaculture* 479:696-703.
- Xing S., Sun R., Pan X., Ma J., Zhang W., Mai K., 2016 Effect of dietary carbohydrates to lipid ratio on growth, body composition, digestive enzyme activities and hepatic enzyme activities in juvenile large yellow croaker, *Larimichthys crocea*. *The Journal of World Aquaculture Society* 47(2):297-307.
- Yamada R., 1983 Pond production system: feed and feeding practice in warmwater fish pond. A state of the art review. Oregon State University, Oregon, pp. 117-144.
- Yanto H., Setiadi A. E., Kurniasih D., 2019 [Influence of the different levels of carbohydrate diets on the growth performance of Tengadak (*Barbonymus schwanenfeldii*)]. *Jurnal Ruaya* 7(2):39-46. [In Indonesian].
- Zhou C., Ge X. P., Liu B., Xie J., Miao L. H., 2013 Effect of high dietary carbohydrates on the growth performance, physiological responses of wuchang bream (*Megalobrama amblycephala*). *Asian-Australasian Journal of Animal Sciences* 26(11):1598-1608.
- Zhou C., Ge X., Niu J., Lin H., Huang Z., Tan X., 2015 Effect of dietary carbohydrate levels on growth performance, body composition, intestinal and hepatic enzyme activities and growth hormone gene expression of juvenile golden pompano (*Trachinotus ovatus*). *Aquaculture* 437:390-397.
- Zhou P., Wang M., Xie F., Deng D. F., Zhou Q., 2016 Effects of dietary carbohydrate to lipid ratios on growth performance, digestive enzyme and hepatic carbohydrate

metabolic enzyme activities of large yellow croaker (*Larmichthys crocea*).  
Aquaculture 452:45-51.

\*\*\* National Research Council (NRC), 2011 Nutrient requirements of fish and shrimp  
Animal Nutrition Series, National Research Council of the National Academies. The  
National Academies Press, Washington D. C., USA, 376 p.

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